**CETACEAN NECROPSY REPORT**

ID Number: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Location: \_\_\_\_\_\_\_\_\_\_ (Lat;Long)\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Reported by: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_ Contact info:\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Date found: \_\_\_\_\_\_\_\_\_\_\_\_\_Date recovered: \_\_\_\_\_\_\_\_\_\_Necropsy Date: \_\_\_\_\_

Species: Age: Sex: M F Unknown

Prosectors: Contact info: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Weather info:\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Human Interaction Forms? 🞎; Chain of custody forms: 🞎; Level A forms filled out 🞎 **BRIEF HISTORY:**

*-Have there been reports of vessel strike in the past week? -What active fisheries are present in the area, and what gear type is used? Where on the whale’s body could this interaction have occurred?*

**GROSS DIAGNOSES:**

**MINIMAL MEASUREMENTS** (cm unless indicated)

|  |  |  |  |
| --- | --- | --- | --- |
| **Basic Measurements** | **Value**  | **Basic Measurements\*** | **Value**  |
| Total length  |  | Skin thickness  |  |
| Girth at axilla (armpit level) |  | Blubber thickness, dorsal bdf **(w/o skin)** straight down from caudal end of dorsal ridge, 45° from spine. |  |
| Girth, anus |  | Blubber thickness, dorsal midline Axillary **(w/o skin)** |  |
| Fluke width |  | Blubber thickness, ventral midline Axillary **(w/o skin)** |  |
|  |  | Blubber thickness, lateral Axillary, **(w/o skin)** |  |

### EXTERNAL EXAMINATION (CIRCLE)

|  |  |  |
| --- | --- | --- |
| **CARCASS CLASSIFICATION:** |  | **Body Condition:** |
| Code 2 Fresh  |  | 1 Robust |
| Code 2.5 mild decomposition  |  | 2 Good |
| Code 3 moderate (decomposed organs intact) |  | 3 Average |
| Code 4 Poor (advance decomposition)  |  | 4 Poor |
| Code 5 Mummified |  | 5 Emaciated |

**GROSS NECROPSY FINDINGS:**

|  |
| --- |
| **PHYSICAL EXAM**1. Photos – made notes on the whale outlines

 Photograph Right side \_\_\_\_Left side \_\_\_\_\_Fluke photo-ID\_\_\_\_\_\_\_ Take photos of any previous tag sites \_\_\_\_\_\_\_\_\_\_  Dorsal if possible \_\_\_\_\_\_\_\_\_Abnormalities\_\_\_\_\_\_\_\_\_\_\_ Photo needs at minimum: ID; measuring device; Date; Site: and which lesion if it is a lesion1. Morphometrics Minimum is: Length:\_\_\_\_\_\_\_\_\_\_\_\_\_Ventral blubber depth at axillary girth;\_\_\_\_\_\_\_\_\_\_\_\_Lateral depth\_\_\_
2. Collect any gear
3. Collect aqueous 18g needle and 20cc syringe
4. Collect Skin sample - zip
5. Barnacles, lice – zip
6. If enough people, collect the eye – Zip
7. Ear plug
 |
| **SQ:** Do the blubber depths above1. Collect a 6 in cubed skin to muscle or larger axillary region DAX \_\_\_\_\_LAX \_\_\_\_\_ or DAX\_\_\_\_\_\_ Another site – Behind the dorsal fin (BDF) \_\_\_\_\_\_\_\_zip
2. Cut parallel lines 12 inches apart and flense. If little time, go for the windows over the bladder / colon; liver/stomach for HABs for and back of head for trauma.
3. Note any discoloration; softness and investigate those areas more.

SQ notes: |
| **MUSCULOSKELETAL (color of muscle, appearance of joint fluid:** Observe muscle under blubber. If any is darker color, or mushier, or liquid, dissect below here to look for broken bones. Collect broken bones for enforcement.  |
| **BODY CAVITIES (FLUID?):** mediastinal, omental, perirenal, cardiac fat? Collect pleural or pericardial fluid if not contaminated.  |
| **RESPIRATORY** (foam, fluid, texture and color of lungs, parasites? – don’t forget the sinuses and blowhole). Is the fat band at the back of the lungs thick?  |
| **CARDIOVASCULAR:** Collect whole blood; examine all valves if possible.Blood sites are ventral peduncle, behind eyes, penis after amputating tip. |
| **ENDOCRINE:** Collect, measure, weigh if possibleAdrenals: Thyroid:  |
| **URINARY:** Collect kidney; Collect urine from bladder (use the window sheet) or Urine \_\_\_\_\_\_ Insert 6 ft. long plastic tube into urethra (cut off the end of the penis in males) **KIDNEY –** Collect for subsampling |
| **LIVER:** bile from ducts; parasites, Note color, texture.Collect liver for subsampling. |
| **DIGESTIVE:** (serosal surface, content, mucosal surface, parasites): Open stomachs – buckets or zips or conical tubes to collectDescribe intestinal contents: Feces – conical tubes or Nalgene bottlesFind and collect Colon LN (Just dorsal to colon caudal towards the Exit) |
| **REPRODUCTIVE:** put 3” pieces of testis, penis, uterus, whole ovary, in Ziploc. Collect entire cervix if possible, or, photograph |
| **SPECIAL SENSES:** Collect aqueous humor from one eye; then Collect eyes – Whirl-pak 18 oz x 2Collect ear plugs and into 15% NBF |
| **NERVOUS SYTEM:** Brain: CSF:Spinal Cord:Brachial Plexus: |

\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

**CARCASS DISPOSITION:**

**LIST SAMPLES SUBMITTED IMMEDIATELY AND WHERE**

**ANCILLARY DIAGNOSTICS:**

Photos 🞎 note who took the photo?

**COMMENTS (CAUSE OF DEATH- WHAT DO YOU THINK HAPPENED?):**

# HUMAN INTERACTION SAMPLES: BULLETS / FISHING GEAR / OTHER:

**HOW DID YOU TAKE YOUR TOXICOLOGY SAMPLES?**

Circle: Ziplocs / Foil / Acetone-cleaned Foil / Teflon / Whirlpak / I-Chem jars

Other\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_ Rinsed tissues with: \_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_

Type of gloves (circle): latex vinyl powder-free nitrile

**SUB-SAMPLING (*asap upon return*):** Keep all samples cool not frozen, subsample into clean containers.

Once sub-sampled and repackaged, label each container with

***Animal ID #; date; sample type***. If possible, DOUBLE Label

**Fluids**:

1. Aqueous humor: Place 1ml aliquots in cryovials and 2 5mls.
2. Blood: Centrifuge whole blood from 2 tubes, separate serum, place in 1 ml aliquots in cryovials (ONLY If super fresh)
3. Urine: Place 5 ml cryovials
4. Feces: Place in clean Whirlpak or better, Centrifuge tubes in 100g aliquots.
5. Bile – Bile: - distribute to an Amber vial and 5 ml cryovials
6. Pleural and pericardial fluids - 5 ml cryovials

Freeze all fluids -70 C if possible, otherwise -20 C

**Tissues**:

1. Skin: 1 piece 1 mm cube into DMSO vial (for genetics)
2. Lesions, skin, lung, heart, kidney, spleen, liver, tongue, intestine, lymph nodes, repro tract, muscle:
	1. 1 cm cube in 10% formalin (all in 1 jar, ensure 10 times as much formalin as total tissue)
	2. 1 cm cube in Whirlpak frozen at -70 C ideal, -20 C OK
3. Bones: Freeze large pieces at -20 C. Small splinters add to formalin jar above
4. Ear plug: Place in jar 15 % formalin, change to 10% formalin after 1 week
5. Blubber: Cut clean cube 4” wide from internal sample, wrap in foil, place in Ziploc, freeze -20 C
6. Parasites: Indicate site collected; Place in 70 % ethanol, Store at room temp.
7. Make sure there is enough formalin on the fixed tissues – if not, trim and add more formalin / bigger containers.

For more detail, follow checklists.

Separate samples for: Biotoxins, Disease, Toxicology, Life history. See checklists



Short lines on whale are Blubber depth sites



The bars indicate sites of blubber thickness measurement



The bars indicate sites of blubber thickness measurement





